

Research Note

Occurrence of arbuscular mycorrhizal fungi in different cropping systems at Cochabamba, Bolivia

Mauritz Vestberg

Agricultural Research Centre of Finland, Plant Production Research, Laukaa Research and Elite Plant Station, Antinniementie 1, FIN-41330 Vihtavuori, Finland, e-mail: mauritz.vestberg@mtt.fi

Marcelo Cardoso and Anna Mårtensson

Swedish University of Agricultural Sciences, Department of Soil Science, PO Box 7014, SE-750 07 Uppsala, Sweden

The occurrence of arbuscule-forming fungi in different cropping systems was investigated in the province of Cercado, Bolivia. The cropping systems included grain and mixed pasture systems, with or without fertilization and agrochemicals. Geographically, the soils were situated at 17°23'9" southern latitude and 66°9'35" western longitude a mean height of 2600 m above sea level. Spores of four arbuscular mycorrhiza fungi-forming genera were observed; *Glomus* Tul. & Tul., *Entrophospora* Ames & Schneider, *Sclerocystis* Berk. & Broome emend. Almeida & Schenck and *Scutellospora* Walker & Sanders. *Glomus* was the dominating genus, followed by *Sclerocystis*; *Scutellospora* and *Entrophospora* were observed occasionally. A cropping system consisting of a native pasture without any fertilization or other plant or soil treatments had the highest numbers of spores and the highest species richness, eight out of nine species identified. The mycorrhizal diversity measured with the Shannon-Wiener index did however not differ very much between cropping systems.

Key words: AMF spore densities, arbuscular mycorrhizal fungi, cropping systems, diversity index, Glomales

Introduction

Arbuscular mycorrhizal fungi (AMF) are amongst the most commonly occurring soil fungi (Gerdemann 1968) and are associated with about 80% of terrestrial plants in most vegetation types (Gianinazzi and Gianinazzi-Pearson 1986). Benefits of the mycorrhiza includes par-

ticularly its contribution to nutrient uptake which has been studied extensively (Hayman 1983, Abbott et al. 1984, Thompson 1990). AMF fungal symbioses, however, not only increase plant nutrient uptake by extending the apparent soil volume available to the plants but improve the tolerance of plants to various biotic and abiotic factors, including pathogens (Gerdemann 1968, Jaizme-Vega et al. 1997) and physical stresses

such as high salt concentrations (Rosendahl and Rosendahl 1991) and drought (Goicoechea et al. 1997).

The arbuscular mycorrhizal fungi (AMF) form a unique order, the Glomales (Morton and Benny 1990), consisting of the six genera, i.e. *Acaulospora* Gerd. & Trappe, *Entrophospora* Ames & Schneider, *Gigaspora* Gerd. & Trappe, *Glomus* Tul. & Tul., *Sclerocystis* Berch & Broome emend. Almeida & Schenck and *Scutellospora* Walker & Sanders. Surveys of species belonging to the Glomales have been conducted in tropical (Redhead 1977, Al-Garni and Daft 1990, Ganesan et al. 1991, Sieverding 1991), temperate (Gerdemann and Trappe 1974, Hall 1977, Walker et al. 1982) and arctic regions (Allen et al. 1987, Väre et al. 1997).

Although data are available on the occurrence of AMF in agricultural systems in eastern Europe, North America and Australia, there is little information concerning agricultural systems of other temperate regions. Most studies of the Glomales in South America have been conducted under tropical conditions (Sieverding 1990, Sieverding 1991, Cuenca et al. 1998). The purpose of this study was to collect, identify and culture species of the Glomales from a field experiment with different cropping systems at Cochabamba, Bolivia. To the authors' knowledge, this is the first report on species of the Glomales from Bolivia.

Material and methods

Study site

Soil samples for estimation of AMF spore densities and establishment of pure cultures of AMF were collected from six fields and a natural ecosystem belonging to the University of Mayor de San Simon (UMSS) in the area of La Tamborada, Cochabamba town, province of Cercado, Bolivia. The latitude and longitude of the sampling area are 17°23'09"S and 66°09'35"W, respectively, and the mean altitude is 2600 m

above sea level. The area has a dry, temperate climate with a mean annual rainfall and temperature of 460 mm and 17°C, respectively. The soil samples were collected between 4 and 27 May 1994. Soil samples from the top soil (0–25 cm) were collected along a diagonal line over the field. Each sample consisted of at least ten individual samples which were pooled and thoroughly mixed to form the final sample of one litre. Six soil samples were collected in a similar manner from the natural vegetation area. The soil samples were stored for two months in a refrigerator (2–3°C) prior to estimation of spore numbers or establishment of AMF trap cultures.

Cropping systems

The UMSS experimental area at La Tamborada included 22 fields with a total area of 117.3 ha. Two fields with low-input cropping systems and three fields with high-input cropping systems were chosen from this area. Two natural ecosystems, a native pasture and a eucalyptus dominant wild ecosystem, were chosen for reference. The cropping systems were not replicated; they occurred only in one place. The size of the individual fields varied between 2.6 and 12.6 ha (Table 1). The experimental areas were all alkaline with pH (in H₂O) varying between 7.4 and 8.6. The phosphorus (P) contents of the soils, estimated as NaHCO₃-extractable P (Olsen et al. 1954) were the highest, 45 and 32 ppm, respectively, in cropping systems 7 (high-input grain/pasture) and 1 (native pasture), and the lowest in the natural ecosystem. The soil content of organic C was also the highest in cropping systems 1 and 7, 3.07% and 1.22%, respectively. The soil of each cropping system had a rather similar texture (fractions of sand, silt and clay), with the exception of field number 4 where the soil had a high content of clay (Table 2).

Spore extraction

Spores were extracted from field soil by wet sieving and decanting (Gerdemann and Nicolson

Table 1. Presentation of cropping systems at Cochabamba, Bolivia.

Cropping system				Size of field, ha
Number	Type	Rotation	Use of fertilizers and pesticides	
1	Natural ecosystem	Native pasture	None	2.5
2	Natural ecosystem	Natural vegetation, <i>Eucalyptus</i> dominating	None	Unknown
3	Low-input grain/pasture	1989–1991, maize 1992, oats 1993, alfalfa	Unknown	3.6
4	Low-input grain	1980–1994, maize	Very low amounts of animal manure, urea and P fertilizers Very low amounts of herbicides used	6.6
5	High-input grain/pasture	1984–1990, alfalfa 1991–1994, maize	Animal manure (amounts unknown) Hormone herbicides used	9.9
6	High-input grain	1980–1994, maize	Heavy applications of animal manure	12.6
7	High-input grain/pasture	1977–1981, grass and alfalfa 1982–1993, maize 1994, oats	Maize fertilized with approx. 100 kg P and urea-N/year Oats given 9–10 tons wet weight of green manure/ha	5.1

1963) followed by centrifugation in water and in a 50% sucrose solution (Walker et al. 1982). A 1000- μm and a 100- μm sieve were used for wet sieving. After centrifugation the spores were

transferred on a filter paper for examination under the dissecting microscope at magnifications up to 50 times with illumination by incident light from a fibre-optic, quartz-halogen light source

Table 2. Chemical and physical properties of soil collected from different cropping systems at Cochabamba, Bolivia. For details of cropping systems, see Table 1.

Cropping system nr	Sand %	Silt %	Clay %	Texture ¹⁾	pH (H ₂ O)	P ²⁾ mg/kg soil	C %
1	32	32	36	CL	7.4	32.0	3.07
2	28	42	30	CL	7.5	4.0	0.15
3	49	22	29	SaCL	8.2	8.4	0.33
4	10	16	74	C	8.6	15.0	0.15
5	50	22	28	SaCL	7.4	22.0	0.88
6	38	24	38	CL	7.6	10.0	0.61
7	32	34	34	CL	7.9	45.0	1.22

¹⁾ Texture: Sand (Sa), Clay (C), Silt (S), Loam (L)

²⁾ Determined as NaHCO_3 -extractable P (Olsen et al. 1954)

with a colour temperature of 3200 K (Walker et al. 1993). Spores were characterized and, when possible, identified to species using a high-power light microscope. Spores from trap cultures and pure cultures were extracted by wet sieving (500- and 50- μm sieves) and decanting and placed in a dish of water for examination under dissecting microscope and a high-power microscope. Mycorrhizal fungal diversity was calculated using the Shannon-Wiener index, which combines two components of diversity, i.e. species richness and evenness of individuals among the species (Krebs 1985).

Fungal isolation and culturing

Pot cultures for trapping AM fungi were established in October 1994 at the Department of Soil Sciences, Uppsala, Sweden. Seeds of *Tagetes* sp. were sown in a 1:1 dilution of original soil and sterilized sand. The pots were kept in a growth chamber under artificial light (16 h light, 260 $\mu\text{mol m}^{-2} \text{s}^{-1}$) at 18°C and 80% relative humidity. The trap cultures were checked for AM fungal growth by wet sieving in March 1995. Newly formed, similar looking spores of the Glomales were then used to establish multi-spore cultures (at the Laukaa Research and Elite Plant Station, Finland) in sealed transparent plastic bags (Sunbag®), incorporating a microfilter to allow gaseous exchange (Walker and Vestberg 1994). The cultures were initiated by placing approximately 20–30 spores directly on the root of young seedlings of *Plantago lanceolata* L. A mixture of steamsterilized sand and perlite (9:1) fertilized with 2 g l⁻¹ bone meal and given 5 g l⁻¹ Dolomite lime, was used as growth substrate. The pure culture were kept at 18/15°C (day/night), 50–60% relative humidity in a growth chamber with warm white artificial light (approx. 120 $\mu\text{mol m}^{-2} \text{s}^{-1}$).

Details of original collection and isolation, resultant cultures and subcultures, and herbarium specimens were recorded in a database developed by C. Walker (Walker and Vestberg 1998). According to that database, each culture

pot was given an 'Attempt' number (unique to each culture attempt made from any original sample) and culture number (sequential for subcultures from a particular attempt). Voucher specimens of fresh material from both trap cultures and from pure cultures were assessed as semi-permanent microscopic slides either in polyvinyl alcohol lacto-glycerol (PVLG) (Omar et al. 1979) or in PVLG with Melzer's reagent (5:1 v/v), (Walker et al. 1993) in the personal herbarium of the first author, each individual collection being given an accession number.

Results

Spore density

Total spore densities varied between 95 per 100 gram dry soil in the low-input cropping system 3 (maize, oats and alfalfa rotation) and 710 in the native pasture (cropping system 1) (Fig. 1a). Nine spore types of AMF belonging to three genera were extracted (Table 3). The genus *Glomus* occurred the most, 90% of all spores extracted, the figures for *Sclerocystis* and *Scutellospora* being only 9.8% and 0.2%, respectively. *Acaulospora* Gerdemann & Trappe, *Gigaspora* Gerdemann & Trappe emend. Walker & Sanders and *Entrophospora* were not found. Three species, *Glomus constrictum* Trappe (Figs. 2 and 3), *G. mosseae* (Nicol. & Gerd.) Gerdemann & Trappe (Fig. 4) and a *Sclerocystis* sp. corresponding in most characteristics to *S. liquidambaris* Wu & Chen (Fig. 5), were found in soil from all cropping systems. Species richness was the highest in the native pasture with eight out of nine spore types. The number of spore types found in the other cropping systems varied between five and six (Table 3). In contrast to the total number of AMF spores and the number of species identified, the Shannon-Wiener diversity index differed little between cropping systems (Fig. 1b).

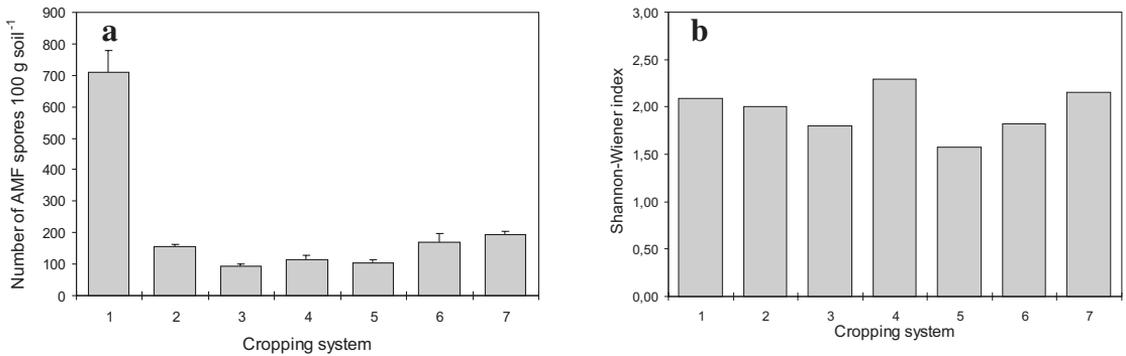


Fig. 1. Spore densities (a) and diversity index as measured by the Shannon-Wiener index (b) of arbuscular mycorrhizal fungi extracted in seven cropping systems (see Table 1) at Cochabamba, Bolivia. Bars represent standard deviations which, however, are only indicative of experimental error.

Pure cultures of Bolivian AMF

Trap cultures with *Tagetes* sp. yielded a *G. microcarpum* Tul. & Tul.-like species (Fig. 6), *G. mosseae*, *G. sp.* "small white", *G. sp.* "shiny brown" and *S. liquidambaris*-like fungus, and these were used to initiate pure cultures. Culturing was successful except with the *G. microcarpum*-like fungus and with the *S. liquidambaris*-like fungus. A commonly occurring *Glomus* sp. "shiny brown" proved to be *G. constrictum*, of

which much darker spores were also found frequently by spore extraction. The *Glomus* sp. called "small white" remained unidentified. One of the cultures was first thought to be a pure culture of *Glomus mosseae* but appeared later to contain also the fungus *Entrophospora infrequens* (Hall) Ames & Sneid. (Fig. 7). Attempts to culture *E. infrequens* alone failed, however. The resulting pure cultures were included in the AMF culture collection of the first author.

Table 3. Number of AMF spores (extracted by wet sieving, centrifugation and sugar floatation) in soil from different cropping systems at Cochabamba, Bolivia. For details of cropping systems, see Table 1.

Fungus	Numbers of AMF spores/ 100 g dry soil							Sum
	Cropping system							
	1	2	3	4	5	6	7	
<i>Glomus constrictum</i>	236	38	6	11	26	65	12	394
<i>G. hoi-aggregatum</i> -like	2	15	2	2		3	12	36
<i>G. fasciculatum</i> -like	30		25	6	4	37	27	137
<i>G. microaggregatum</i> -like	2				2			4
<i>G. microcarpum</i> -like	2							2
<i>G. mosseae</i>	110	17	25	18	33	28	22	253
<i>G. sp.</i> "greyish"	143			4			2	149
<i>Sclerocystis liquidambarum</i> -like	48	24	4	8	2	7	2	95
<i>Scutellospora</i> sp.		1						1

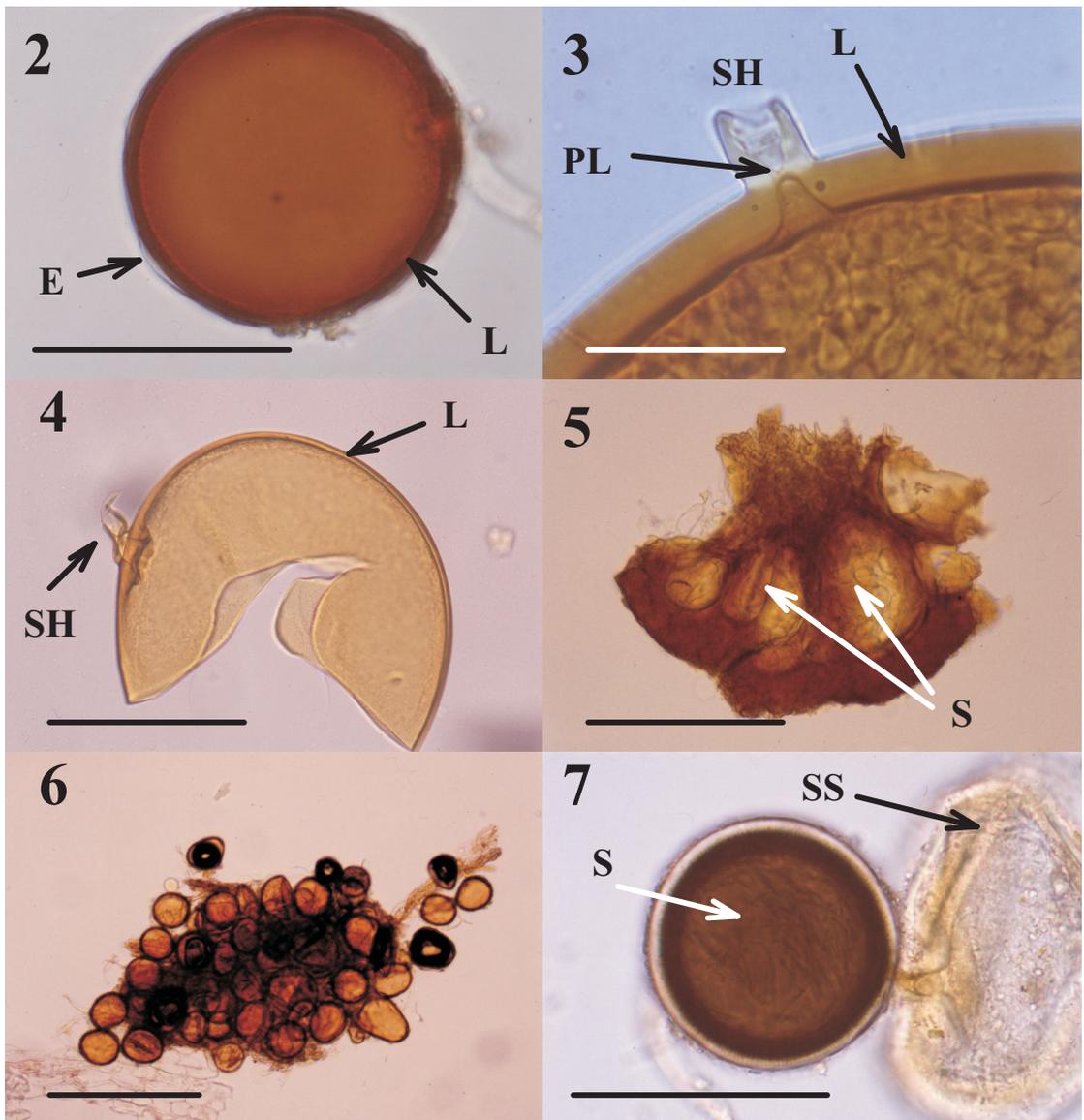


Fig. 2. Intact spore of *Glomus constrictum* showing a dark brown laminated wall component (L) and remnants of a hyaline outermost wall component (E). From pure culture attempt 775-1, bar 100 μm .
 Fig. 3. Part of a spore of *Glomus constrictum* showing a laminated wall component (L), a short subtending hypha (SH) and a plug (PL) formed in it. From pure culture attempt 775-1, bar 30 μm .
 Fig. 4. Crushed ectocarpic spore of *Glomus mosseae* from soil of cropping system 5 showing the funnel-like subtending hypha (SH) and a relatively thin laminated wall component (L); bar 200 μm .
 Fig. 5. Part of a sporocarp of *Sclerocystis liquidambaris* showing individual spores (S). From cropping system 5; bar 200 μm .
 Fig. 6. Sporocarp-like dense cluster of a *Glomus* sp. resembling *G. microcarpum*. From soil of cropping system 3; bar 300 μm .
 Fig. 7. Spore (S) and sporiferous saccule (SS) of *Entrophospora infrequens* found in pure culture attempt 773-2, bar 100 μm .

Discussion

Spores of the genus *Glomus* were found most frequently followed by the genus *Sclerocystis* which was represented only by a species resembling *S. liquidambaris* (Wu and Chen 1987). The genus *Scutellospora* was found only once in the native ecosystem while the genera *Acaulospora* and *Gigaspora* were not detected at all. This result partly agrees with the survey of locations for new species descriptions by Allen (1991). According to this survey, new species of *Glomus* have been described in all climatic regions, but especially in the temperate regions, while species of the genera *Acaulospora* and *Sclerocystis* have been most frequently in the tropics. The absence of *Acaulospora* in this study might have been partly due to the use of too big sieves. Some small spores of *Acaulospora* will pass through a 100- μm sieve. A better sieve size would have been 50 μm . On the other hand, small spores easily stick to soil particles and to each other which means that spores measuring 70–80 μm in diameter can also be detected when using a 100 μm sieve. Repeated trapping might have revealed more slowly sporulating *Acaulospora* and *Scutellospora* as was found by Stutz and Morton (1996).

Species diversity was highest in the native pasture with eight out of nine AMF spore types found, while the other cropping systems exhibited 5–6 spore types out of nine. Similar results have been obtained in other investigations. Douds et al (1995) found that soil in low-input agriculture had greater populations of *Glomus occultum* Walker type spores and other *Glomus* spp., whereas the conventionally farmed soil had greater populations of *G. etunicatum* Becker & Gerdemann-type spores. Another study (Douds et al. 1993) showed higher spore numbers of *Gigaspora gigantea* (Nicol. & Gerd.) Gerdemann & Trappe in low-input plots than in conventional plots. The fungus resembling *S. liquidambaris* was found in all cropping systems in our study despite different levels of fertilizer application and a pH exceeding 7.0. This result disagrees

with the study of Sieverding (1990), according to which liming and fertilization of native tropical systems quickly made all the *Sclerocystis* spp. disappear.

The spore densities were the highest in the native pasture without any fertilizer or pesticide inputs, but much lower in the other low-input and high-input cropping systems as well as in the natural ecosystem. This result is in agreement with findings by Sieverding (1991), according to whom AMF spore densities are generally the highest low-input agricultural sites while the numbers are often lower at both native and high-input sites. Schenck and Siqueira (1987) also found more spores per unit of soil in agroecosystems than in native ecosystems, but a greater species diversity in the native ecosystems than in the agroecosystems. Douds et al. (1993) found higher populations of AMF spores in low-input plots than in conventionally farmed plots. Sattelmacher et al. (1991) found higher AMF root colonization in rye growing in an organic biological-dynamic farming system than in a conventionally managed high-input farming system.

The spore densities in this study must, however, be regarded only as indicative of real differences between the cropping systems because of the lack of replication of cropping systems across soil type. The standing crops also differed between systems. Maize was the standing crop in systems 4, 5 and 6, while oats and alfalfa were growing in system 7 and 3, respectively. Systems 1 and 2 had a mixture of plant species. Differences in host plants have been shown to result in differential AMF sporulation in the field (Kruckelmann 1975, Schenck and Kinloch 1980, McGraw and Hendrix 1984, Koske 1987, Janke and Peters 1993).

Spore densities of different cropping systems seemed not to correlate with differences in soil P which varied considerably, from 4 to 45 mg P g soil⁻¹. This result disagrees with the wellknown fact that increasing amounts of phosphorus has a negative impact on mycorrhiza (Antunes and Cardoso 1991, Arias et al. 1991, Bolan 1991, Fairchild and Miller 1990, Hung et al. 1990, Raju et al. 1990). Mårtensson and Carlgrén (1993)

found that the number of mycorrhizal spores decreased rapidly over time with increasing annual additions of a soluble phosphorus fertilizer. In our study there are indications that the amount of organic carbon in the soil better explains the differences in spore densities. The native pasture which had the highest spore density also had the highest content of organic carbon, 3.07%, followed by cropping system 7 with 1.22%. The rest of the cropping systems had soil with amounts of organic C below 1%. A positive cor-

relation between soil organic matter and mycorrhiza has been found also in other investigations (Saif 1986, Sieverding 1991, Toro and Sieverding 1986).

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Vestberg, M. et al. Arbuscular mycorrhizal fungi in Bolivia

SELOSTUS

Arbuskelimykorrhizasientien esiintyminen eri viljelyjärjestelmissä Cochabambassa, Boliviassa

Mauritz Vestberg, Marcelo Cardoso ja Anna Mårtensson
Maatalouden tutkimuskeskus ja Ruotsin maatalousyliopisto

Tutkimuksessa selvitettiin arbuskelimykorrhizasientien (AMS) esiintymistä Mayor de San Simon -yliopiston viljelyjärjestelmäkokeessa Cochabamban kaupungissa, Boliviassa. Alue sijaitsee n. 2600 metrin korkeudessa ja siellä vallitsee kuivahko lauhkea ilmasto. Maanäytteitä kerättiin toukokuussa 1994 viideltä peltolohkolta, yhdeltä luonnonlaitumelta ja yhdeltä viljelemättömältä alueelta, jossa oli eukalyptuspuiden alla luontaista kasvillisuutta. Maanäytteistä laskettiin AMS-itiöiden määrät ja sienet pyrittiin luokittelemaan myös lajin mukaan. AM-sieniä pyydystettiin maasta myös *Tagetes*-kasvin avulla. Joistakin sienilajeista perustettiin puhdasviljelmiä heinä-

ratamon juuristoon.

Tutkimuksessa selvitettiin ensimmäisen kerran AM-sientien esiintymistä Boliviassa. Alueelta löydettiin neljään sukuun kuuluvia sienilajeja; *Glomus*, *Entrophospora*, *Sclerocystis* ja *Scutellospora*. *Glomus*-sukuun kuuluvia sienilajeja määritettiin ylivoimaisesti eniten (90 %). Luonnonlaitumella maassa oli yli kolme kertaa enemmän AMS-itiöitä kuin eri tavalla lannoitetuissa peltolohkoissa, joissa viljeltiin maisia, palkokasveja ja heinäkasveja. Myös viljelemätön luonnonkasvillisuuden alue sisälsi vähemmän AMS-itiöitä kuin luonnonlaidun.

