

The effect of ammonium ferric hexacyanoferrate on reducing radiocaesium transfer from grass silage to sheep

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A study was carried out to examine the effect of ammonium ferric hexacyanoferrate (AFCF) on the transfer of radiocaesium from grass silage to the tissues of male lambs. During ensiling, a formic acid based additive and AFCF were sprayed on grass contaminated with ^{134}Cs and the mixture was allowed to incubate for 45 days. A dose of $21 \text{ mg AFCF d}^{-1}$, fed to sheep offered contaminated silage for fourteen days, reduced ^{134}Cs transfer to muscle by 45% compared to that of control sheep. An equivalent dose of AFCF administered in a capsule reduced transfer by only 3%. In another experiment, AFCF intake of 50, 100 and 150 mg d^{-1} for ten days reduced ^{134}Cs transfer to sheep muscle by 75, 82 and 86%, respectively. In control lambs, of average live weight 38 and 47 kg, the feed to muscle ^{134}Cs transfer coefficient averaged 0.15 d kg^{-1} , but equilibrium between tissue and feed ^{134}Cs had probably not been reached due to the short feeding period. Increasing doses of AFCF from 0 to 150 mg d^{-1} increased the faecal/urinary ^{134}Cs ratio from 2 to 42.

Key words: AFCF, excretion, preservation, radiocaesium, sheep, silage, transfer

Introduction

Different materials have been administered to ruminants for reducing radiocaesium transfer to meat and milk products. These include bentonite (Andersson 1989, Beresford et al. 1989,

Mitchell et al. 1989), zeolite (Phillippo et al. 1988, Unsworth et al. 1989), vermiculite (Hazard 1969), kaolin (Giese 1989), stable caesium (Oughton et al. 1991) and crude fibre (Johnson et al. 1968). Organic complexes such as ferric hexacyanoferrates, commonly referred to as

Paasikallio, A. et al. Effect of AFCF on reducing ¹³⁴Cs transfer from grass silage to sheep

Prussian Blue (PB) compounds are, however, the most effective caesium binders on a unit weight basis (Giese 1989, Unsworth et al. 1989). PB compounds reduce the intestinal absorption of radiocaesium, and are most effective when administered with contaminated feed. PB compounds react with radiocaesium in the gut to form a complex that is excreted in the faeces. Furthermore, they have been reported to enhance tissue excretion of radiocaesium into the gut (Hove et al. 1990, Nielsen et al. 1990, Åhman 1996). Ammonium ferric hexacyanoferrate (AFCF) is the most commonly used PB compound and binds radiocaesium in the gut in exchange for an ammonium ion. AFCF has been used as a feed supplement in silage and concentrates for housed ruminants and in salt licks and boli for grazing animals (Unsworth et al. 1989, Pearce et al. 1989, Hove et al. 1990, Voigt 1993, Hansen et al. 1996, IAEA 1997). In general, studies have only considered the effects of AFCF mixed with silage some time prior to feeding (Arnaud et al. 1988, Giese 1989, Unsworth et al. 1989, Vreman et al. 1992), while information concerning the use of AFCF at grass harvesting and its effects during and after storage in silo is limited.

In the 1960s, studies were carried out on the toxicity of PB on animals and humans (Nigrovic 1963, Madhus et al. 1966). After radioactive emissions following the Chernobyl accident, toxicological as well as other studies on PB were revived (Arnaud et al. 1988, Giese 1988, Nielsen et al. 1990, Pearce 1994). The results showed that the consumption of milk and meat from PB treated animals could be considered safe with respect to human health. So far, the application of AFCF to soil directly or via the faeces does not seem to have any negative effects on soil-plant environments (Vandenhove et al. 1997, 1998, 2000). However, long-term and more comprehensive studies are lacking (Jones et al. 1999).

Following the Chernobyl accident, PB compounds were officially approved for use as a feed additive in Russia, Ukraine, Belarus, Norway, Germany and Austria (IAEA 1997). The

EC directive allows the use of AFCF as a feed additive in the EU countries, but its use should be authorized at the national level (Commission Directive 1996). PB compounds have also been used successfully with human casualties of the Goiânia radiation accident in Brazil (Lipsztein et al. 1991, Melo et al. 1994).

This study was carried out to examine the effect of low levels of AFCF, mixed in contaminated grass during ensiling, on the transfer of ¹³⁴Cs to ovine tissues. The hypothesis tested was that, in situations of radioactive fallout, AFCF could be mixed with contaminated grass at harvesting together with an acid based ensiling additive.

Material and methods

Preparing ¹³⁴Cs-AFCF-silage

Ryegrass was grown in pots in peat soil contaminated with ¹³⁴Cs. Grass was harvested after 45 days. One kilogram of contaminated ryegrass was placed on a plastic sheet and thoroughly mixed with 6 kg of uncontaminated chopped and prewilted timothy-meadow fescue grass. The purpose of prewilting was to minimize silage effluent production. Grass was treated with a formic acid based additive (AIV2-solution containing 80% formic acid and 2% orthophosphoric acid) at a rate of 5 ml kg⁻¹. Additive diluted with water (50%) was sprayed on grass prior to ensiling. In addition, AFCF (ammonium ferric hexacyanoferrate, NH₄Fe³⁺[Fe²⁺(CN)₆], containing 33% NH₄Cl) as a water solution was also applied to grass. AFCF was applied at a rate of 0 and 100 mg kg⁻¹ and 0, 250, 500 and 750 mg kg⁻¹ of grass in experiments 1 and 2, respectively. Once applied, grass (7 kg) was ensiled in laboratory silos (volume of cylinder 15.4 dm³, diameter 14 cm). Each silo was fitted with a drainage system for collecting and monitoring silage effluent production. After filling, each silo was tightly sealed and weighted with con-

Table 1. Some aspects of experimental conditions (daily AFCF doses, ^{134}Cs intake and sheep parameters).

	Experiment 1	Experiment 2
AFCF doses ($\text{mg d}^{-1} \text{ sheep}^{-1}$)	0, 21 ^a , 21	0, 50, 100, 150
^{134}Cs intake ($\text{kBq d}^{-1} \text{ sheep}^{-1}$)	7	13
Test period (d)	14	10
Number of replicates	2	3
Total number of sheep	6	12
Age of sheep (d)	171	214
Live weight (kg)	38	47

^a AFCF (ammonium ferric hexacyanoferrate) given in capsule, others in silage

crete blocks and water bags to a pressure of 585 kg m^{-2} . Preservation lasted for about 45 days. After opening the silos, silage was sampled and divided into daily doses (0.2 kg/sheep) and frozen in plastic bags for later use. Fermentation quality of both ^{134}Cs contaminated and uncontaminated silages was good.

Animals and experimental design

Male crossbred Finnish Landrace lambs, mean age 171 (experiment 1) and 214 days (experiment 2) and live weight of 38 and 47 kg, respectively, were used in the study (Table 1). Lambs were allocated according to live weight into two blocks of three animals (experiment 1) and into three blocks of four animals (experiment 2). Lambs within each block were exposed to different experimental treatments. The animals were individually housed in metabolic cages. Before the experiments started, the animals were allowed 7 days to become accustomed to the cages. In experiment 1, silage contaminated with ^{134}Cs was given daily to all sheep for fourteen days. AFCF was given either with contaminated silage or in gelatine capsules. Daily intakes of AFCF were 0, 21 and 21 mg d^{-1} for control, treated silages and AFCF capsules (two

sheep each), respectively. In experiment 2, contaminated silage with different AFCF doses was given to twelve sheep for ten days. The intake of AFCF in silage were 0, 50, 100 and 150 mg d^{-1} for three sheep each (Table 1). Experiments 1 and 2 were conducted in September 1995 and October 1996, respectively.

Feeding and sampling

In both experiments, the sheep received a daily ration fed in the mornings, consisting of 3 kg of uncontaminated farm silage, 0.3 kg of barley and 0.2 kg of ^{134}Cs contaminated silage (with or without AFCF). When AFCF (21 mg d^{-1}) was given in a capsule, the capsule was administered immediately after the morning feed (experiment 1). The daily feed intake of each animal was recorded in both experiments. Water was available *ad libitum* with consumption recorded for all experimental animals. Urine and faeces were collected separately and output was measured daily. Sub-samples of urine and faeces were stored frozen prior to ^{134}Cs determinations. At the end of both experiments, the animals were slaughtered and sampled. In experiment 1, dorsal and femoral muscle, heart, liver, kidney, whole blood and digesta samples were col-

lected. For the second experiment, samples of neck and femoral muscle, diaphragm, heart, liver and kidney were collected. In all cases, samples were stored frozen and subsequently monitored for ¹³⁴Cs.

Sample analysis

For activity measurement animal tissues were cut into small pieces. Visible fat was removed from samples of muscle and kidney. The ¹³⁴Cs activity concentration of contaminated grass silage, urine, faeces and animal tissues was determined using a low-background semiconductor spectrometer coupled to a germanium detector. Activity concentrations of ¹³⁴Cs are presented on a fresh weight (FW) basis. Transfer coefficients of ¹³⁴Cs (d kg⁻¹) from feed to tissues was calculated as ¹³⁴Cs activity concentration in tissues (Bq kg⁻¹) per ¹³⁴Cs intake (Bq d⁻¹).

Statistical methods

In both experiments, the effect of AFCF treatments on tissue ¹³⁴Cs activity concentration was evaluated by Analysis of Variance for repeated measures using the MIXED PROCEDURE within SAS (SAS 1992). Repeated measurements within each animal were found to be highly correlated, a factor which was taken into account by using a compound symmetry covariance structure assigned on the basis of Akaike's and Schwarz's Bayesian information criteria (Wolfinger 1996). The statistical model (Gumperetz and Brownie 1993) used to assess the effect of treatments was:

$$Y_{ijk} = \mu + A_i + B_j + e_{ij} + T_k + (AT)_{jk} + (BT)_{ik} + h_{ijk}$$

where Y_{ijk} is the observed response (e.g. ¹³⁴Cs activity concentration), μ is the intercept, A_i the fixed effect from the i th treatment, B_j the random block effect. e_{ij} is a random effect that represents the error associated with the ij th cell. T_k and $(AT)_{jk}$ represent the fixed effect of tissue

and treatment-tissue interactions, respectively, while $(BT)_{ik}$ is the random effect of block-tissue interaction and h_{ijk} are error terms.

Due to a low activity concentration in blood (experiment 1), the effect of different AFCF treatments on ¹³⁴Cs activity concentration in blood was analyzed separately by Analysis of Variance for a randomized complete block design as follows:

$$Y_{ij} = \mu + A_i + B_j + e_{ij}$$

where Y_{ij} , μ , A_i , B_j and e_{ij} are equivalent to those in the previously described model.

Differences between treatments and between tissues were tested using orthogonal contrasts. In experiment 2, linear and quadratic effects of AFCF treatment were studied. Prior to analysis, data concerning ¹³⁴Cs activity concentration was transformed into natural logarithms to the constancy of error variance. Assumptions of both models were checked using graphical methods i.e. box-plot for normality and plots of residuals to ensure constancy of error.

Results

Experiment I

The sheep consumed their whole feed ration. When AFCF (21 mg d⁻¹) was administered to sheep as a capsule for fourteen days, the ¹³⁴Cs activity concentration of tissues except the liver did not differ from that of the controls. When an equivalent dose of AFCF was administered in silage, the average transfer of ¹³⁴Cs to tissues was reduced to 50% of that of controls (Table 2). Activity concentrations of ¹³⁴Cs differed significantly ($P < 0.05$) between tissues in controls, being highest in kidneys and lowest in muscle. The feed to muscle transfer coefficient for ¹³⁴Cs in control sheep was twice as high as in animals treated with AFCF administered in silage (Table 3).

AGRICULTURAL AND FOOD SCIENCE IN FINLAND

Vol. 9 (2000): 135–147.

Table 2. Activity concentration (least square mean with 95% confidence interval) and reduction (% of control) of ^{134}Cs in sheep tissues after administering ^{134}Cs contaminated silage daily, without ammonium ferric hexacyanoferrate (AFCF) (control) and with 21 mg d^{-1} of AFCF given as a capsule or in silage fed for 14 days (experiment 1).

Tissue	^{134}Cs in sheep					Significance		
	Control	AFCF 21 mg d^{-1} (capsule)		AFCF 21 mg d^{-1} (silage)		P_1	P_2	P_3
	(Bq kg^{-1})	(Bq kg^{-1})	(%)	(Bq kg^{-1})	(%)			
Muscle	961 (770, 1199)	942 (755, 1176)	3	522 (418, 651)	46	0.89	< 0.005	< 0.005
Heart	1163 (932, 1452)	960 (769, 1198)	17	559 (448, 697)	52	0.18	< 0.005	0.01
Liver	1483 (1189, 1851)	918 (736, 1146)	38	598 (479, 746)	60	0.01	< 0.001	0.02
Kidney	2032 (1628, 2536)	1773 (1420, 2212)	13	1011 (810, 1262)	50	0.33	< 0.005	< 0.005
Whole blood	127 (47, 339)	84 (31, 225)	34	54 (20, 146)	57	0.33	0.12	0.31

Number of sheep per treatment = 2, muscle is a mean of dorsal and femoral muscles, P_1 = control vs. AFCF (capsule), P_2 = control vs. AFCF (silage), P_3 = AFCF (capsule) vs. AFCF (silage)

Table 3. Transfer coefficients of ^{134}Cs (muscle/silage) for sheep administered daily with different doses of ammonium ferric hexacyanoferrate (AFCF).

AFCF dose (mg d^{-1})	n	Test period (d)	^{134}Cs transfer coefficient	
			Mean (d kg^{-1})	SD
<i>Experiment 1</i>				
0	2	14	0.14	0.01
21 ^a	2	14	0.14	<0.01
21	2	14	0.07	0.02
<i>Experiment 2</i>				
0	3	10	0.16	0.03
50	3	10	0.04	0.01
100	3	10	0.03	0.01
150	3	10	0.03	0.01

n = number of sheep, SD = standard deviation

^a AFCF given in capsule, others in silage

Paasikallio, A. et al. Effect of AFCF on reducing ¹³⁴Cs transfer from grass silage to sheep

Table 4. Activity concentration of ¹³⁴Cs in digesta after administering ¹³⁴Cs contaminated silage daily, without ammonium ferric hexacyanoferrate (AFCF) (control) and AFCF given as a capsule or in silage fed for 14 days (experiment 1).

Gastrointestinal site	¹³⁴ Cs in digesta (Bq kg ⁻¹)								
	Control			AFCF (capsule)			AFCF (silage)		
	n	Mean	SD	n	Mean	SD	n	Mean	SD
Rumen	2	403	79	2	254	79	2	575	115
Omasum	2	495	138	1	510	–	1	1626	–
Abomasum	2	391	33	2	406	6	2	877	11
Duodenum	1	408	–	2	388	18			
Jejunum, oral+aboral	2	576	48	2	483	131	2	526	60
L.intestine+caecum	2	1313	26	2	1425	32	1	2356	–
L.intestine, prox.	1	1761	–	2	1805	262	2	2397	129
L.intestine, dist.	1	3279	–				2	4209	1129
L.intestine+rectum				1	3536	–			
Rectum	1	2766	–	1	2462	–			

n = number of sheep, SD = standard deviation, data are given in fresh weight

Gastrointestinal tract contents had higher ¹³⁴Cs activity concentrations in AFCF (silage)-treated sheep than in the other treatment groups. In general, the ¹³⁴Cs level of digesta tended to increase towards the posterior part of gastrointestinal tract being highest in the distal part of the large intestine (Table 4).

Excretion of ¹³⁴Cs expressed as a % of ¹³⁴Cs intake was greater in faeces than urine (Table 5). Urinary ¹³⁴Cs excretion was significantly ($P < 0.001$) higher in sheep fed the control diet. Faecal ¹³⁴Cs excretion increased continuously before reaching a plateau after 5–6 days (Fig. 1).

Experiment 2

During the first day two sheep refused to consume silage. After this, the sheep consumed all given feed. Tissue ¹³⁴Cs activity concentrations were clearly higher in control animals than in those treated with AFCF, therefore controls were not included in further comparisons be-

tween treatments. Caesium 134 activity concentration of tissues except the liver differed significantly ($P < 0.05$) between the three AFCF-treatments. When AFCF was given to sheep at 50, 100 and 150 mg d⁻¹ in silage for ten days, ¹³⁴Cs transfer to muscle reduced by 75, 82 and 86% relative to control values, respectively (Table 6). In control animals, the ¹³⁴Cs activity concentration in kidneys was significantly ($P < 0.001$) higher than that determined in all other tissues. The feed to muscle transfer coefficient of ¹³⁴Cs was 0.16 d kg⁻¹ in controls, while for AFCF-treated sheep, a value around 0.03 d kg⁻¹ was observed (Table 3).

Faecal excretion of ¹³⁴Cs was clearly lower, and urinary excretion higher, in control sheep than in AFCF-treated sheep (Fig. 2). There were, however, only negligible differences in ¹³⁴Cs excretion between AFCF treatments (Table 5). Daily water intake of individual animals varied from 0.07 to 1.81 l d⁻¹. Feed and water intakes were not affected by the AFCF-treatments.

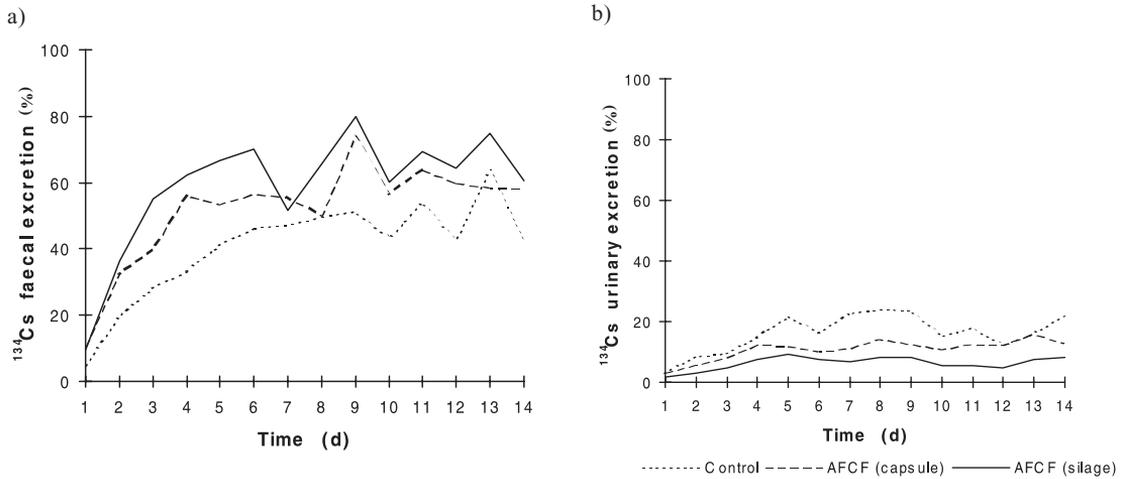


Fig. 1. Effect of ammonium ferric hexacyanoferrate (AFCF) (21 mg d^{-1}) administered in a capsule or in silage on ^{134}Cs faecal (a) and urinary (b) excretion (% of intake) during fourteen days. (experiment 1)

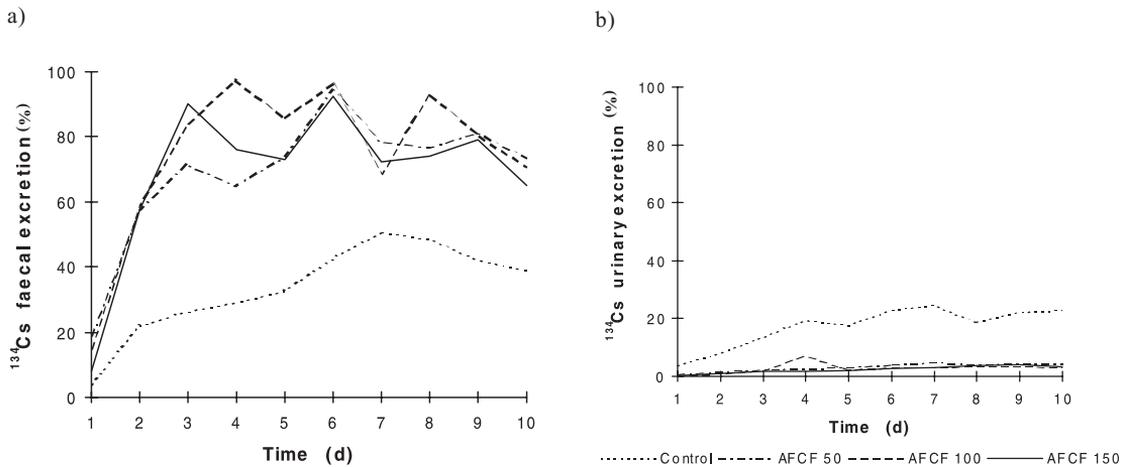


Fig. 2. Effect of different doses of ammonium ferric hexacyanoferrate (AFCF) ($50, 100$ and 150 mg d^{-1}) on ^{134}Cs faecal (a) and urinary (b) excretion (% of intake) during ten days. (experiment 2)

AGRICULTURAL AND FOOD SCIENCE IN FINLAND

Paasikallio, A. et al. Effect of AFCF on reducing ¹³⁴Cs transfer from grass silage to sheep

Table 5. Mean ¹³⁴Cs excretion (% of intake) in faeces and urine and the faecal/urinary ratio (F/U) of sheep administered ¹³⁴Cs contaminated silage daily with different doses of ammonium ferric hexacyanoferrate (AFCF).

AFCF dose (mg d ⁻¹)	n	Test period (d)	¹³⁴ Cs excretion (%)				¹³⁴ Cs F/U
			Faeces		Urine		
			Mean	SD	Mean	SD	
<i>Experiment 1</i>							
0	2	14	41	15	16	6	3
21 ^a	2	14	52	16	11	4	5
21	2	14	59	18	6	3	10
<i>Experiment 2</i>							
0	3	10	34	16	17	8	2
50	3	10	69	21	3	1	23
100	3	10	75	28	3	3	33
150	3	10	69	26	2	2	42

n = number of sheep, SD = standard deviation

^a AFCF given in capsule, others in silage

Table 6. Activity concentration (least square mean with 95% confidence interval) and reduction (% of control) of ¹³⁴Cs in sheep tissues after administering ¹³⁴Cs contaminated silage daily, without ammonium ferric hexacyanoferrate (AFCF) (control) and with different doses of AFCF given in silage for 10 days (experiment 2).

Tissue	¹³⁴ Cs in sheep						Significance		
	Control	AFCF 50 mg d ⁻¹	%	AFCF 100 mg d ⁻¹	%	AFCF 150 mg d ⁻¹	%	P ₁	P ₂
	(Bq kg ⁻¹)	(Bq kg ⁻¹)		(Bq kg ⁻¹)		(Bq kg ⁻¹)			
Muscle	2205 (1662, 2925)	557 (420, 739)	75	393 (296, 521)	82	305 (230, 405)	86	< 0.005	0.78
Heart	2407 (1814, 3193)	620 (467, 823)	74	465 (350, 617)	81	393 (296, 521)	84	0.03	0.72
Liver	2756 (2077, 3657)	641 (483, 851)	77	545 (411, 723)	80	447 (337, 593)	84	0.08	0.92
Kidney	4524 (3410, 6002)	1065 (802, 1413)	76	748 (563, 992)	85	698 (526, 926)	85	0.04	0.41
Diaphragm	2652 (1999, 3519)	682 (514, 905)	74	461 (347, 612)	83	402 (303, 534)	85	0.01	0.45

Number of sheep per treatment = 3, muscle is a mean of neck and femoral muscles, P₁ = linear trend and P₂ = quadratic trend of AFCF levels without control.

Discussion

Reduction of ^{134}Cs

In this study, the lowest AFCF dose of 21 mg d^{-1} fed in silage, reduced the final ^{134}Cs activity concentration in ovine muscle by 46%, which was further decreased by 86% at the highest dose of 150 mg d^{-1} . According to Pearce et al. (1989), a bolus providing AFCF $20\text{--}24 \text{ mg d}^{-1}$ reduced the ^{137}Cs level of sheep muscle by 42% and in animals receiving 200 mg d^{-1} the reduction was 85%. Intakes of AFCF from salt licks or boli of between $25\text{--}300 \text{ mg d}^{-1}$, have reduced the transfer of ^{137}Cs to sheep tissue by 50–90% (Hove 1993). In reindeer, an AFCF dose of 500 mg d^{-1} prevented the absorption of ^{137}Cs almost completely (Åhman 1996). A comparison of the current results with those documented in the literature, indicates that incubation with silage did not reduce the efficacy of AFCF to inhibit radiocaesium transfer to sheep tissues. In this respect, the use of AFCF would be suitable for field conditions. The AFCF doses used in the present study were considerably lower than those generally recommended for small ruminants ($1\text{--}2 \text{ g d}^{-1}$) (Giese 1988, 1989). Silage AFCF contents, as high as 500 mg kg^{-1} , would not exceed recommended AFCF daily doses in sheep fed that silage 3 kg d^{-1} .

In experiment 1, AFCF given in silage, was more effective in reducing tissue ^{134}Cs levels than the same dose of AFCF administered via capsules. Administered in silage, AFCF may have bound ^{134}Cs during ensiling, whilst when AFCF is given in capsule form, ^{134}Cs probably has a greater chance of being transferred to tissues before AFCF is released into the rumen. This hypothesis was supported by digesta activity concentrations, since ^{134}Cs activity concentrations in the rumen-abomasum content of animals receiving AFCF in silage was higher than in capsule fed animals. Radiocaesium activity

concentrations of control sheep was significantly higher in renal than in other tissues, which is in accordance with other studies (Howard and Lindley 1985, Howard et al. 1989).

Transfer coefficients

In control animals, the transfer coefficient for radiocaesium from feed to muscle averaged 0.15 d kg^{-1} . The value was rather low compared, particularly, to those of post-Chernobyl studies (Howard et al. 1987, 1989, Andersson 1989, Beresford et al. 1989, Howard 1989). The low value was probably due to the short feeding periods (10 and 14 days) used in the current study. The transfer coefficients for radiocaesium are valid only when the radiocaesium level of tissue has equilibrated with intake, which in sheep is attained between 20 and 30 days post-contamination (Howard et al. 1989, Pearce et al. 1989).

Excretion of ^{134}Cs

A plateau for faecal ^{134}Cs excretion seems to be reached after 5–6 days from the beginning of the experiment (large day-to-day variation). This was a rather short period compared to the findings of Vandecasteele et al. (1989) who reported a faecal excretion plateau after 20 days in pregnant ewes. In control sheep, faecal ^{134}Cs excretion was twice that in urine. Mean faecal ^{134}Cs excretion accounted for 38% of intake, which was somewhat lower than values (50%) reported by Beresford et al. (1989) and Vandecasteele et al. (1989). However, the contamination period in their studies was much longer (34 and 76 days, respectively) than in the present experiment. The administration of AFCF mixed in silage increased faecal ^{134}Cs excretion considerably in this study, and was attributed to AFCF inhibition of ^{134}Cs intestinal absorption.

Conclusion

Adding AFCF to contaminated silage considerably reduced ¹³⁴Cs transfer to sheep tissues. Incubation in silage did not reduce the efficacy of AFCF as a Cs-binder. In this respect, use of AFCF in the field would be possible. However, additional studies are needed to develop spraying technics for AFCF application during ensiling in combination with formic acid or enzyme

inoculant silage additives under field conditions. Furthermore, long term studies are required to validate ¹³⁴Cs transfer coefficients, since the feeding period was too short to reach equilibrium between tissue and feed ¹³⁴Cs in the current study.

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SELOSTUS

Lihan ¹³⁴Cs-aktiivisuuspitoisuuden vähentäminen ferriheksasyanoferraatin avulla

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Radiocesiumin kulkeutumista märehtijöihin on voitu vähentää syöttämällä eläimille erilaisia cesiumia sitovia mineraaleja kuten esimerkiksi bentoniittia, zeoliittia, vermikuliittia ja kaoliinia sekä kuitupitoista rehua. Ferriheksasyanoferraatit (preussinsininen, PB-yhdisteet) ovat edellisiä huomattavasti tehokkaampia cesiumsitojia. Ammoniumferriheksasyanoferraatti (AFCF) on PB-yhdisteistä tehokkaimpia ja eniten tutkittu. AFCF sitoo itseensä radiocesiumia vaihtaen sen ammoniumioniin. Koska PB ei imeydy ruoansulatuskanavasta kudoksiin, poistuu radiocesium eläimestä PB-yhdisteeseen sitoutuneena sonnan mukana. Paras tulos saavutetaan kun cesiumsitojaa syötetään samanaikaisesti aktiivisen rehun kanssa. Joissakin tapauksissa sitojan on myös todettu poistavan elimistössä jo olevaa radiocesiumia. Sisätiloissa PB:tä tavallisesti syötetään eläimille kerran pari päivässä sekoittamalla sitä pienen määrään rehua. Laitumella sitojaa voidaan helpoimmin antaa lisäämällä sitä nuolukiveen tai bolukseen. Lukuisten tutkimusten perusteella PB-yhdisteitä voidaan pitää vaarattomina eläimille ja ihmisille, sillä yhdisteiden ei ole todettu havaittavissa määrin hajoavan ja imeytyvän kudoksiin ja maitoon.

Tutkimuksen tarkoituksena oli selvittää pienten AFCF-määrien, joita muhitettiin aktiivisessa säilörehussa sen kypsymisen ajan, vaikutusta radiocesiumin kulkeutumiseen lampaan kudoksiin. Muhittamisen vaikutusta cesiumsitojan tehokkuuteen ei ole aikaisemmin selvitetty. Tutkimuksen taustalla oli ajatus, että laskeumatilanteessa AFCF voitaisiin ruiskuttaa, kuten säilöntäainekin, saastuneeseen ruohoon jo pellolla sadonkorjuun yhteydessä.

AFCF:n vaikutusta tutkittiin puolivuotiailla pääsikärisoilla, joita pidettiin aineenvaihduntahäikeissä. Raiheinää kasvatettiin astiakokeissa turvemaassa, johon oli lisätty ¹³⁴Cs:a. Näin saatua aktiivista raiheinää lisättiin silputtuun pellolla kasvatettuun raiheinään, ja seokseen ruiskutettiin säilöntäainetta sekä veteen sekoitettua AFCF:ia. Säilörehun valmistuttua se jaettiin päiväannoksiin. Lampaille syötettiin AFCF:ia päivittäin neljäntoista päivän ajan 21 mg kapselissa ja 21 mg rehussa (koe 1) ja päivittäin kymmenen päi-

vän ajan 50, 100 ja 150 mg rehussa (koe 2). Kontrollilampaat eivät saaneet AFCF:ia.

Kudosten ¹³⁴Cs-aktiivisuuspitoisuus oli yleensä korkein munuaisissa ja matalin lihaksessa. Jo pienetkin AFCF-määrät vähensivät selvästi lampaan lihan ¹³⁴Cs-aktiivisuuspitoisuutta. Rehun mukana annettu päivittäinen 21 mg AFCF-annos vähensi radiocesiumin kulkeutumista lampaan lihakseen 45 % kontrollieläimeen verrattuna. Sama AFCF-määrä annettuna kapselissa vähensi kulkeutumista 3 %. Eläinten saadessa päivittäin aktiivisen rehun mukana 50, 100 ja 150 mg AFCF:ia väheni radiocesiumin kulkeutuminen lihakseen 75, 82 ja 86 %. Tässä tutkimuksessa käytetyt AFCF-määrät olivat pienille märehtijöille suositeltuja annoksia (1–2 g päivässä) huomattavasti pienempiä.

Lihäs/rehu-siirtokertoimet laskettiin jakamalla lihaksen ¹³⁴Cs-aktiivisuuspitoisuus (Bq kg⁻¹) päivittäisen rehuannoksen sisältämällä ¹³⁴Cs-määrällä (Bq d⁻¹). Kokeessa 1 siirtokertoimet olivat 0,14 (kontrolli), 0,14 (kapseli) ja 0,07 d kg⁻¹ (rehu). Kokeessa 2 kontrollieläinten siirtokerroin oli 0,16 ja muiden keskimäärin 0,03 d kg⁻¹. Kontrollieläinten siirtokertoimet olivat pienempiä kuin useissa Tshernobylin jälkeä suoritetuissa tutkimuksissa, minkä katsottiin johtuvan tämän tutkimuksen kokeiden lyhytaikaisuudesta. Eräiden selvitysten mukaan tasapaino kudosten radiocesiumipitoisuuden ja radiocesiumin jatkuvan saannin välillä saavutetaan lampaille vasta noin 20–30 päivän kuluttua.

Radiocesiumin erittyminen sonnan mukana oli kontrollilampaille selvästi vähäisempää ja virtsan mukana runsaampaa kuin AFCF:ia saaneilla lampaille. Kun rehussa annettun AFCF:n päiväannosta nostettiin 0:sta 21 mg:aan, radiocesiumin keskimääräinen erittyminen sonnan mukana oli vastaavasti 41 ja 59 % ja virtsan mukana 16 ja 6 % radiocesiumin saannista (koe 1). Kun AFCF-annosta nostettiin 0:sta 150 mg:aan, vastaavat radiocesiumin erittymismäärät olivat sonnassa 34 ja 69 % ja virtsassa 17 ja 2 % (koe 2).

Tutkimuksessa saatiin alustavaa tietoa aktiivisessa säilörehussa muhineen AFCF:n vaikutuksesta lampaan lihan radiocesiumtasoon. AFCF:n muhitta-

minen useita viikkoja säilörehussa ei vähentänyt sen tehoa cesiumsitojana; pienetkin AFCF-määrät vähensivät huomattavasti radiocesiumin kulkeutumista eläimen kudoksiin. Ainakin tältä osin AFCF:n käyttö pellolla olisi mahdollista. AFCF:n ruiskutustekni-

kan kehittämiseksi tarvitaan lisätutkimuksia. Lisäksi lihas/rehu- siirtokertoimien luotettavuuden tarkistaminen vaatisi pitempiaikaisia ruokintakokeita kuin mitä tässä tutkimuksessa oli mahdollista tehdä.